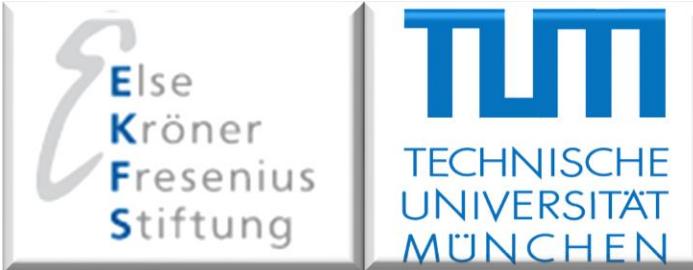




DZHK

DEUTSCHES ZENTRUM FÜR
HERZ-KREISLAUF-FORSCHUNG E.V.

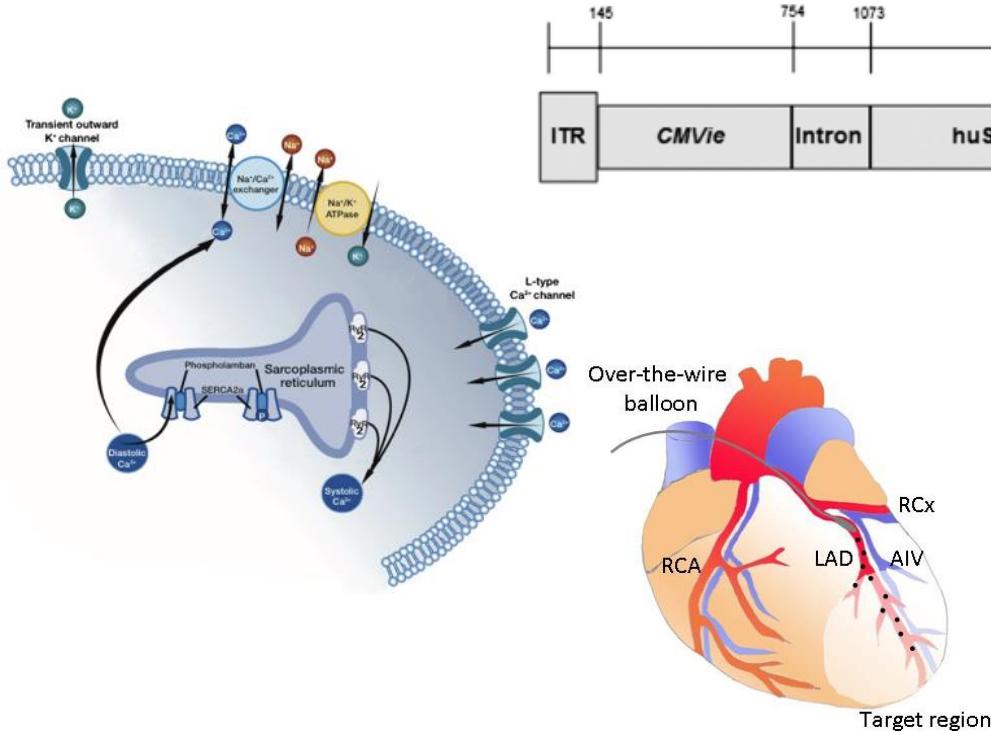


Gene therapy and gene editing – Examples of successful translation

C. Kupatt

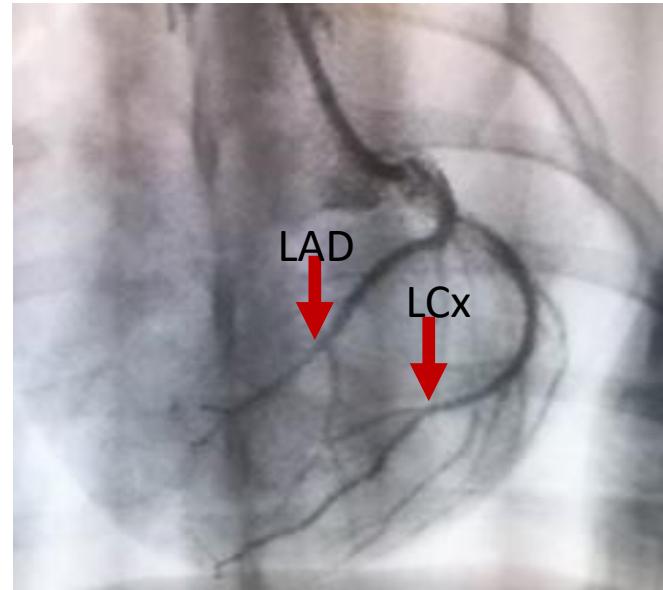
Medizinische Klinik und Poliklinik I
Klinikum rechts der Isar, München

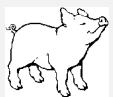
AAV1-Serca as heart failure gene therapy



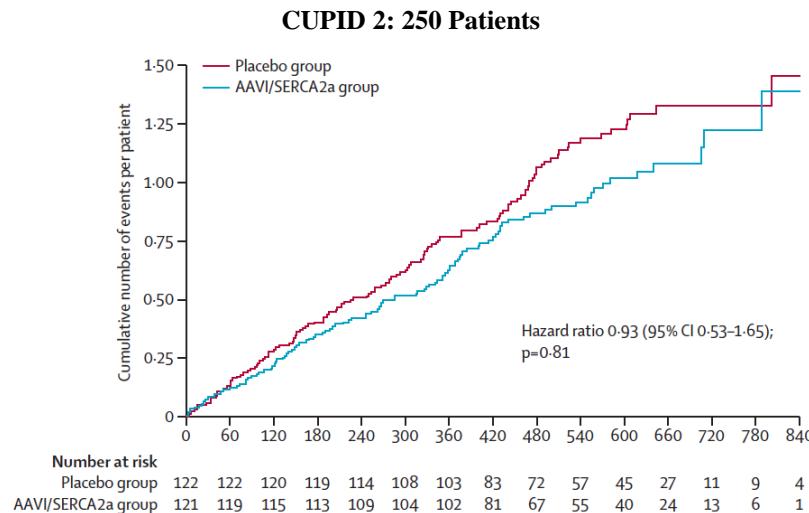
Greenberg et al.,
JACC Heart Failure 2014

● Therapeutic agent
Hinkel & Kupatt, Cardiovasc Res 2012





Phase II Study of AAV1-SERCA2a in Heart Failure

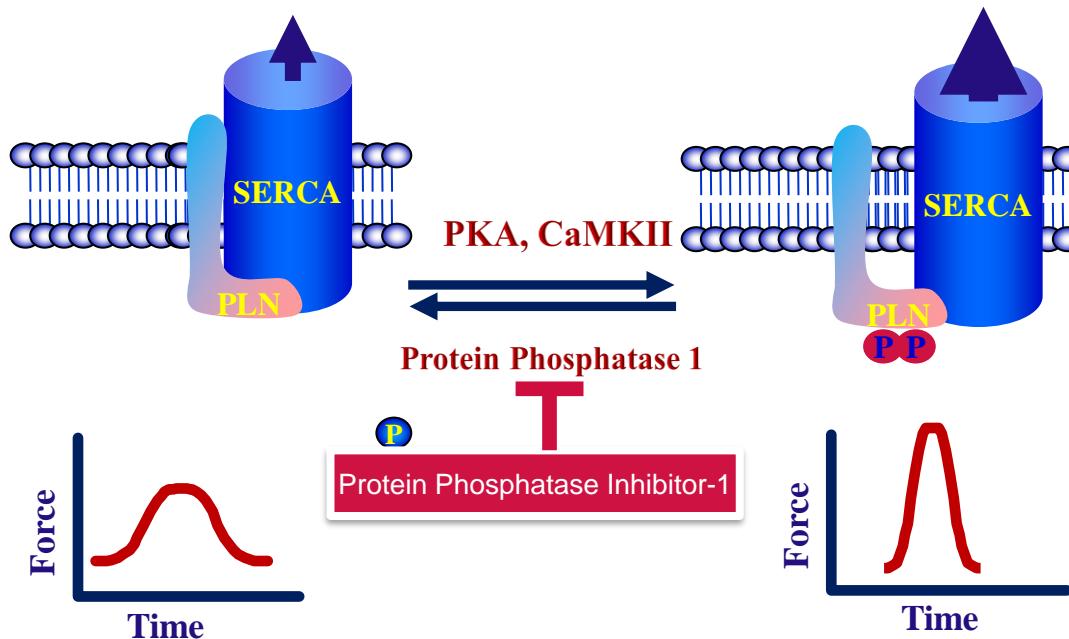


Greenberg et al., Lancet 2016

Viral Uptake in Animal Models of Heart Failure

Animals	Virus	Delivery	ss copies of viral DNA/ μ g DNA	% Infected Cardiac Cells
Mice	AAV9.SERCA2a	Intravenous	42,000	~75%
Rats	AAV9.SERCA2a	Intravenous	30,000	~70%
Pigs	AAV1.SERCA2a	Intracoronary	8,000	~30%
Sheep	AAV6.SERCA2a	Intracoronary	9,000	~33%
	AAV1.SERCA2a	Surgical, MCARD	13,000	~42%
Humans	AAV1.SERCA2a	CUPID AGENT HF SERCA-LVAD	<561 ND <80	1% 0 <0.2%

PP1 Is Regulated by Inhibitor-1



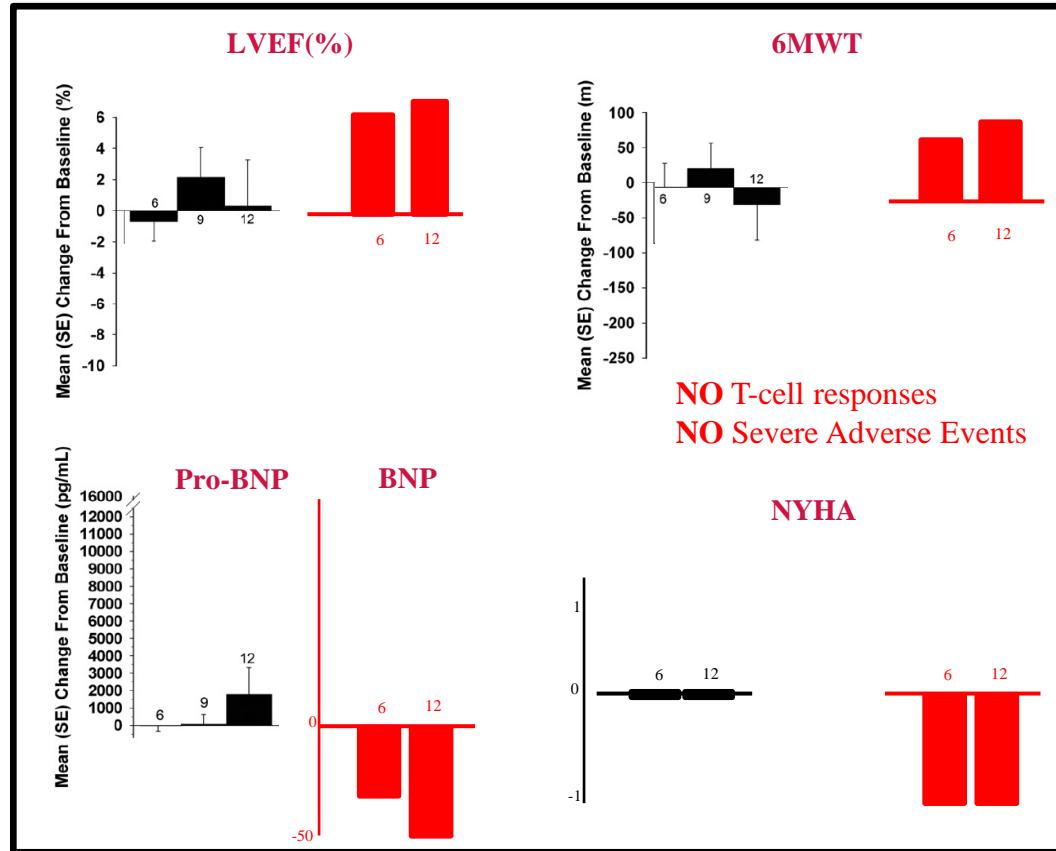
Significant Clinical Efficacy of Dose-Escalation



10^{13} vg/patient in
CUPID Trial by
Celladon

3×10^{13} vg/patient in
Cohort A of MUSIC
Trial by Sardocor

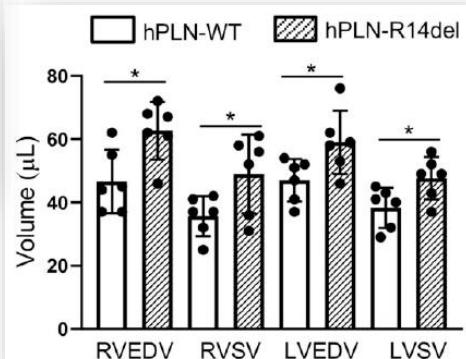
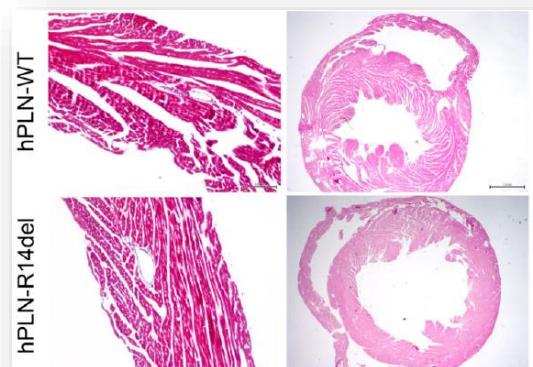
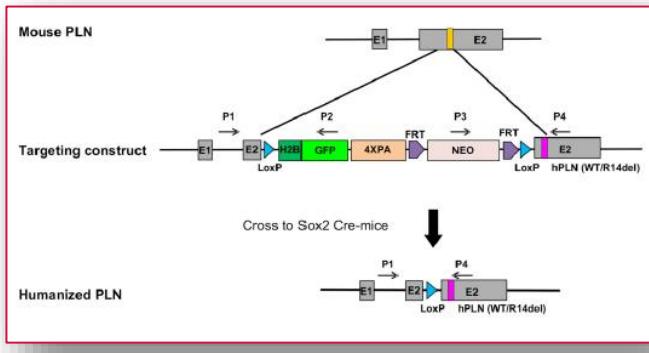
Further escalation in
Cohort B to be
performed in 1Q2023



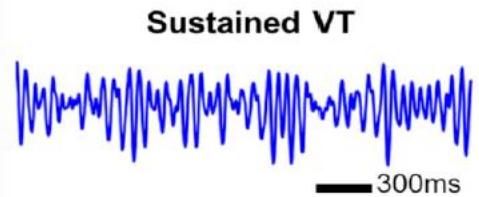
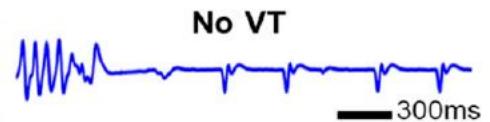
Successfully gene-edited or gene-supplemented genetic cardiomyopathies

- **Hypertrophy**
- *MYH7* ←
- *MYBPC*
- **Dilatation**
- *RBM20*
- *Titin*
- **ARVC / AVC**
- *PKP 2*
- *DMD* ←
- *PLN R14del* ←

Phospholamban R14 del mutation

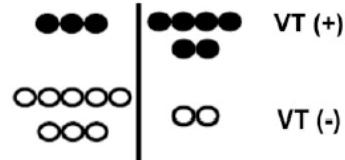


vECG Response to ISO & Rapid Pacing

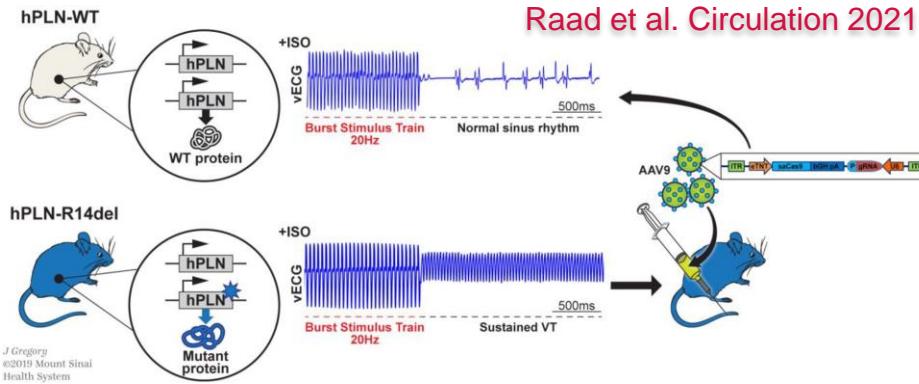
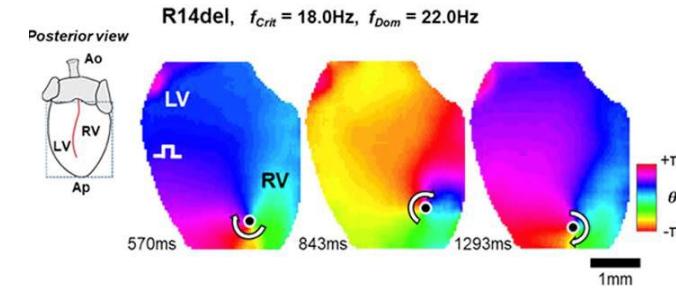
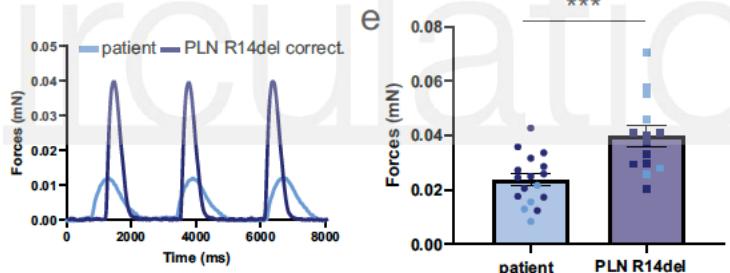
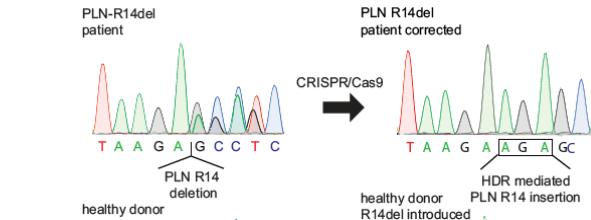


VT inducibility
(up to pre-defined cutoff)

hPLN-WT hPLN-R14del



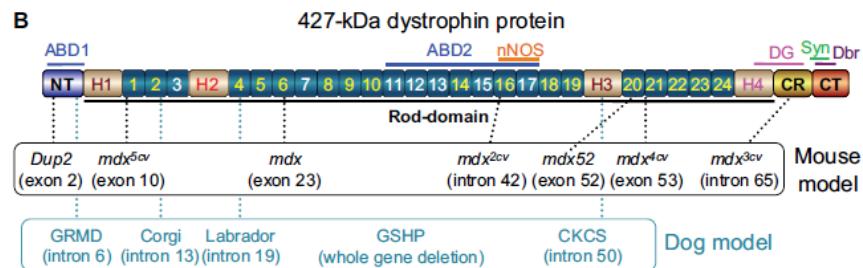
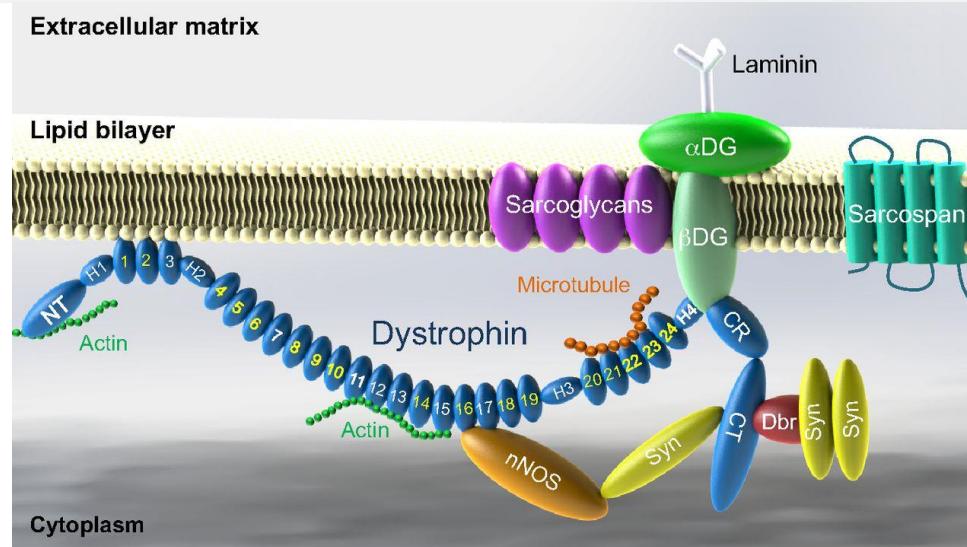
Reversion of PLN R14del arrhythmogenic phenotype by Cas9 editing in vivo (mouse)



Feyen et al., Circulation 2021

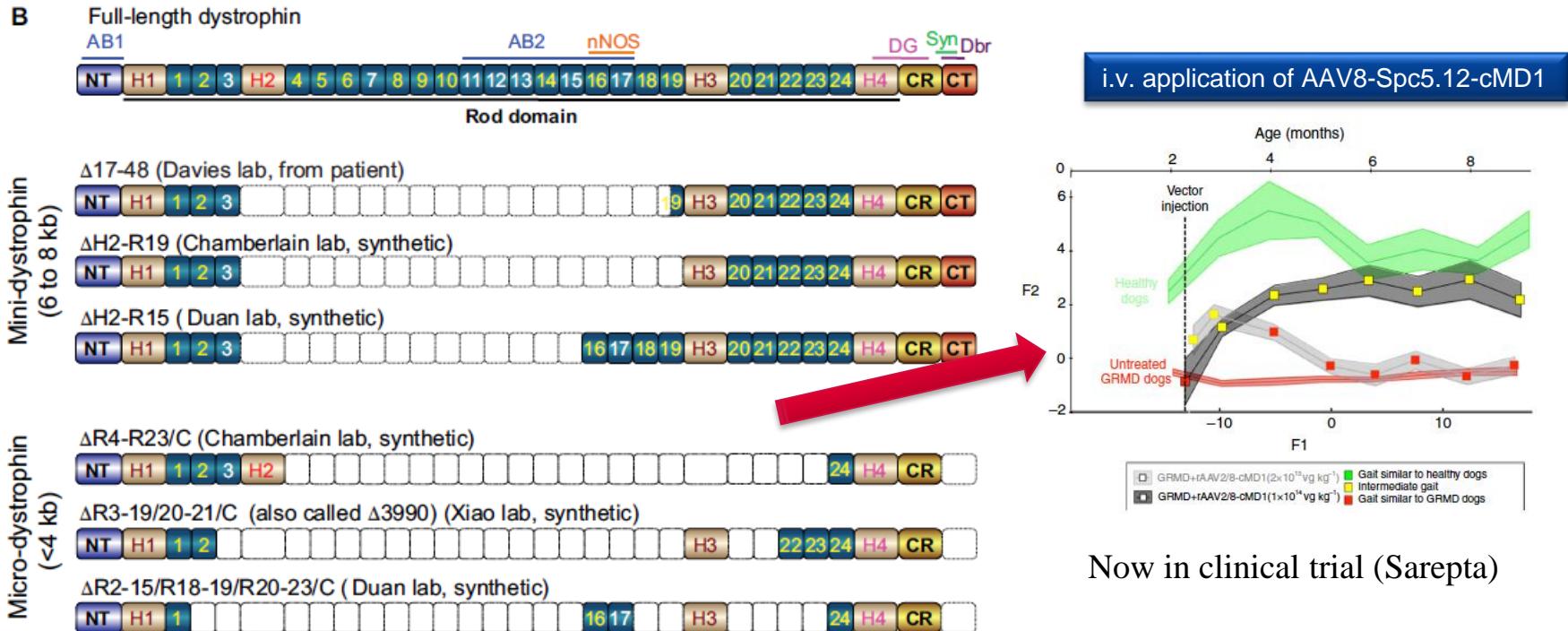
Dave et al., Cardiovasc Research 2022

Dystrophin

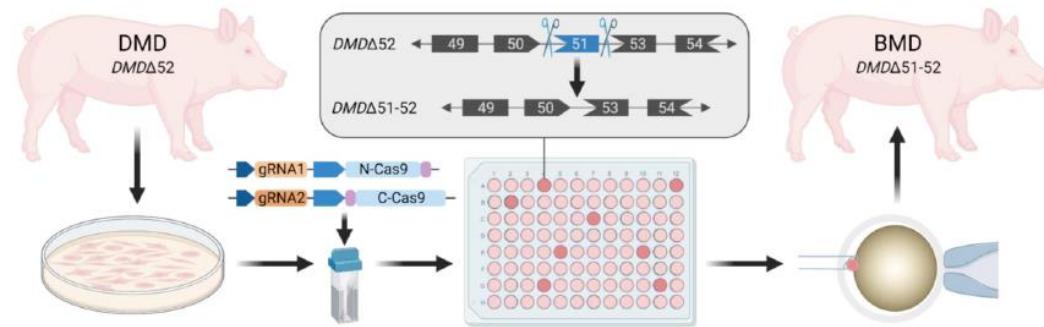


McGreevy et al., 2015

Mini- and microdystrophins



Genotype of DMD and BMD pigs



WT

exon 50 exon 51 exon 52 exon 53

```
••• ATT GAA GCA CCT ACT AGT CAG ••• AAG CAG AAG GCA ACA CTG ••• ACT GAT CGA ATT GAA AGA ATA •••
```

DMD

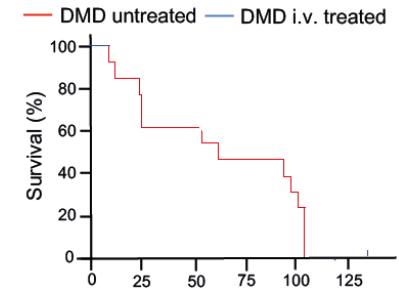
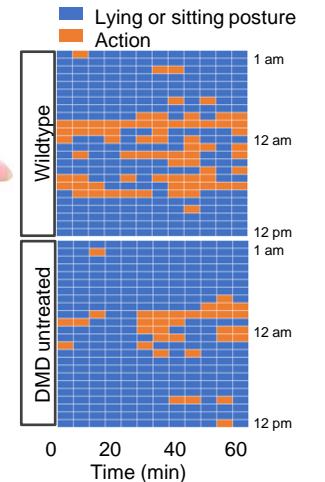
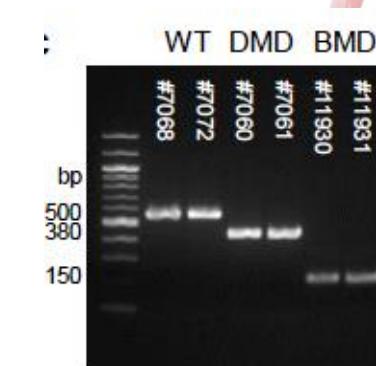
exon 50 exon 51

```
••• ATT GAA GCA CCT ACT AGT CAG ••• AAG CAG AAG
```

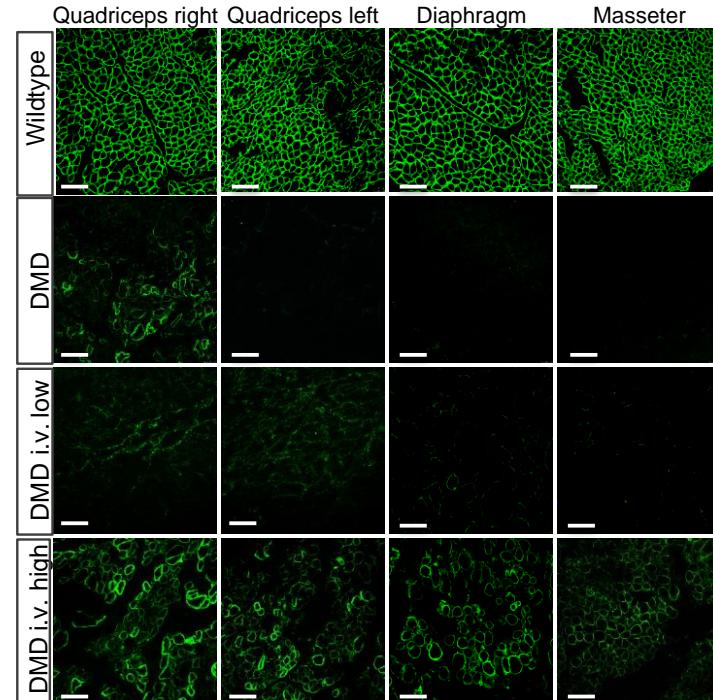
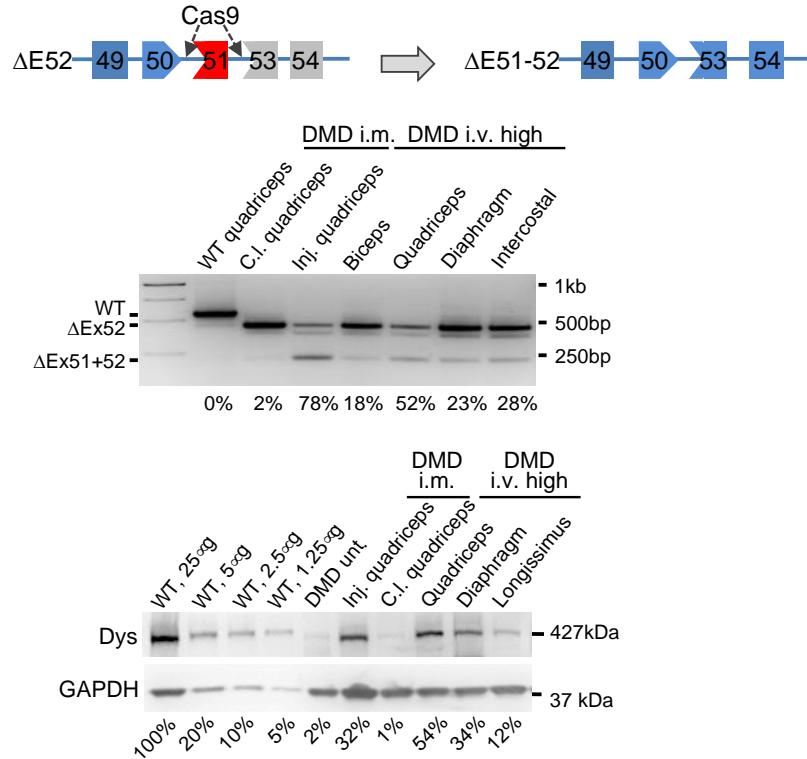
BMD

exon 50

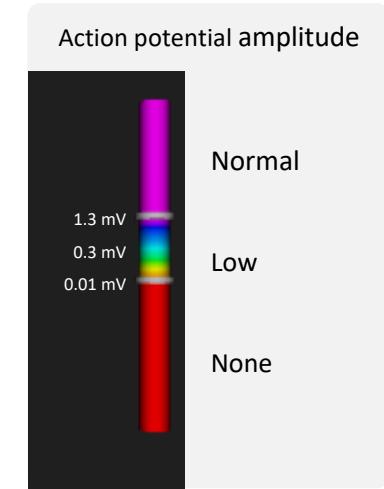
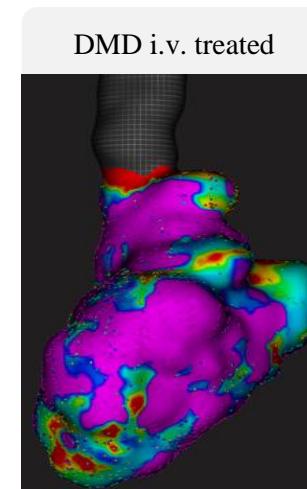
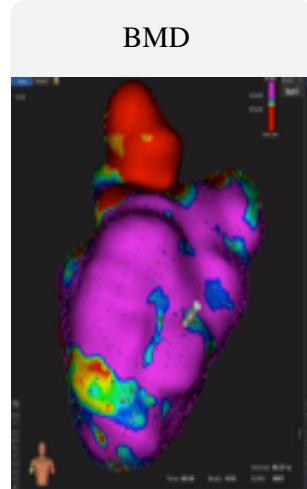
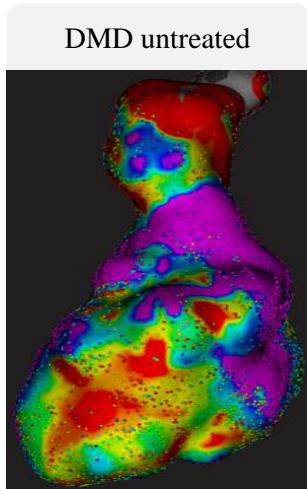
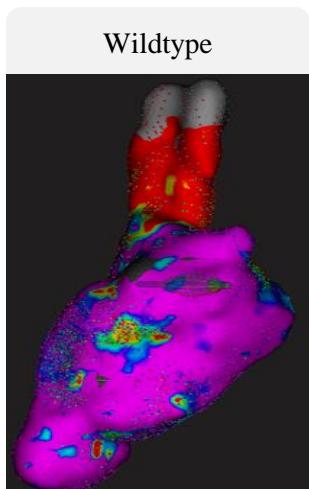
```
••• ATT GAA GCA C
```



DMD pig (Δ 52) treated with AAV-Cas9-gE51

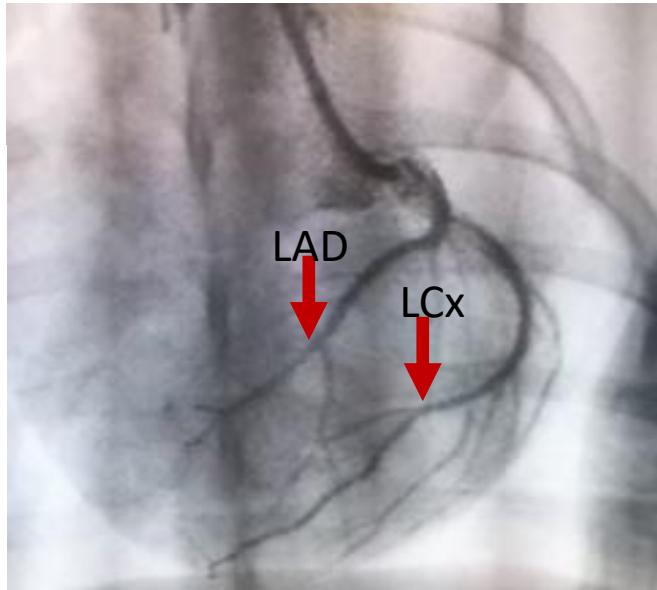
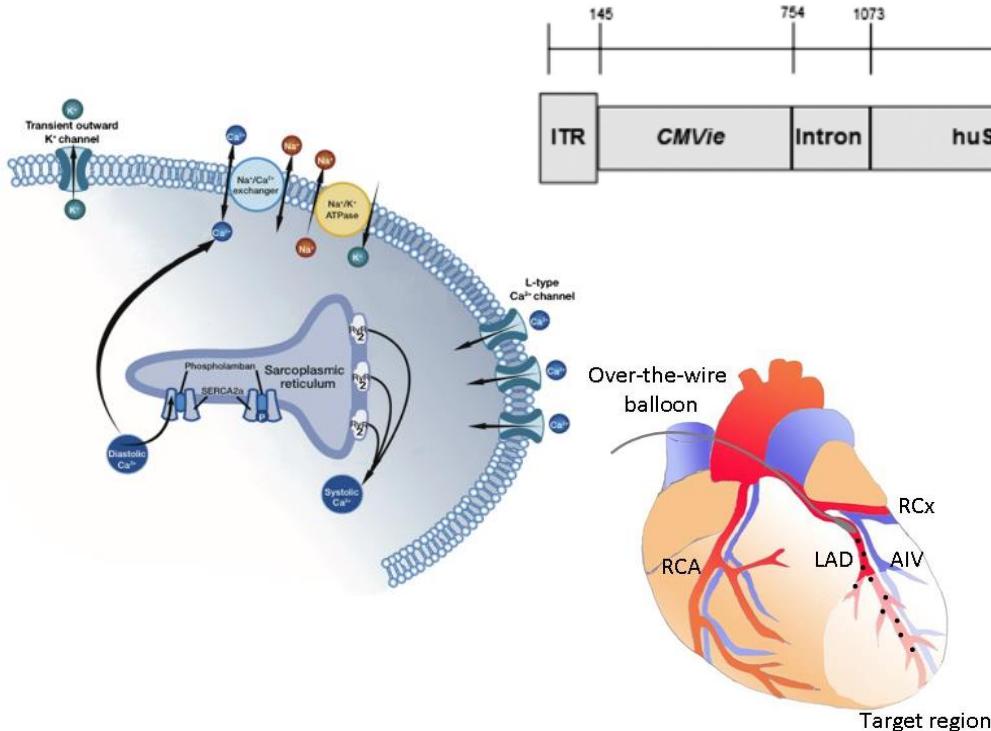


High resolution electrophysiologic analysis of DMD and BMD pigs



Moretti et al., Nat Med
2020

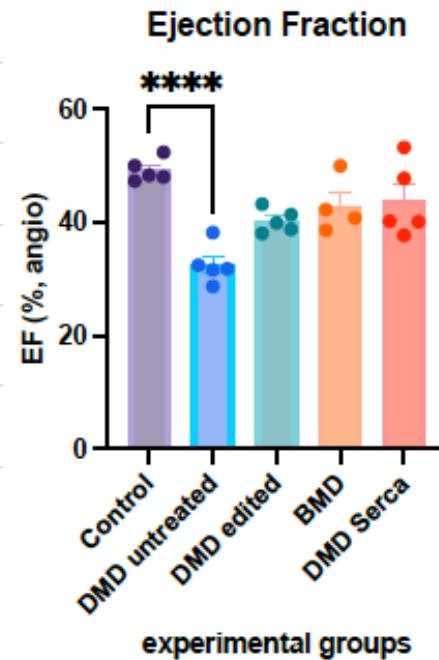
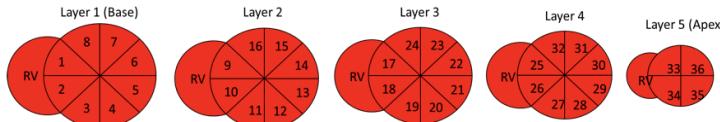
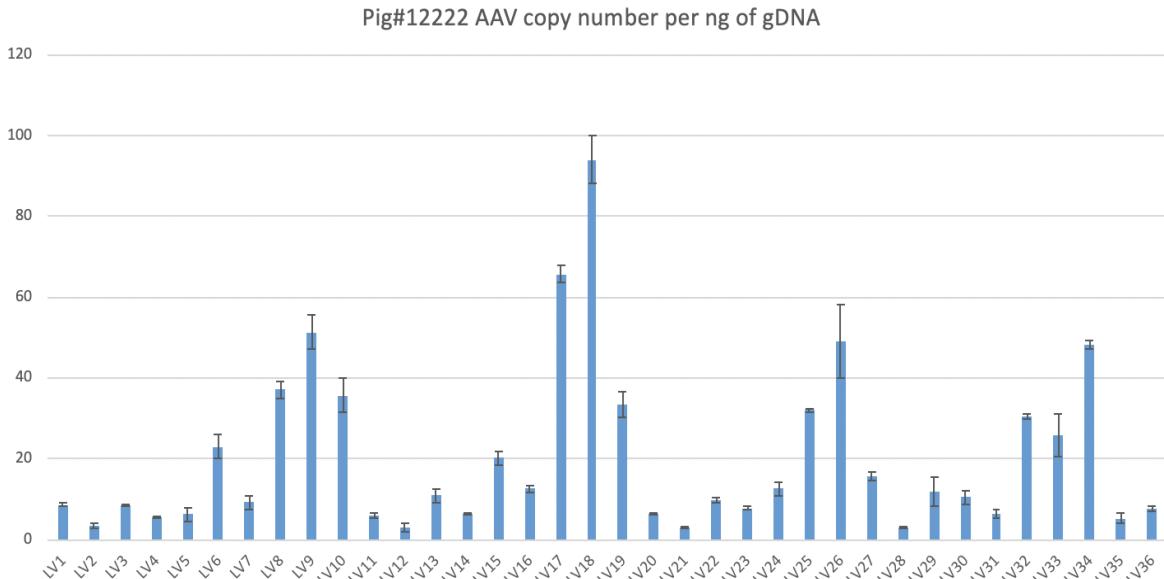
AAV1-Serca as heart failure gene therapy



Greenberg et al.,
JACC Heart Failure 2014

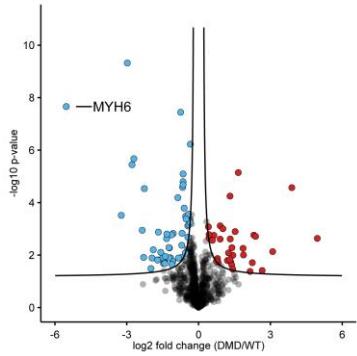
● Therapeutic agent
Hinkel & Kupatt, Cardiovasc Res 2012

Serca2a Expression and function in pig hearts

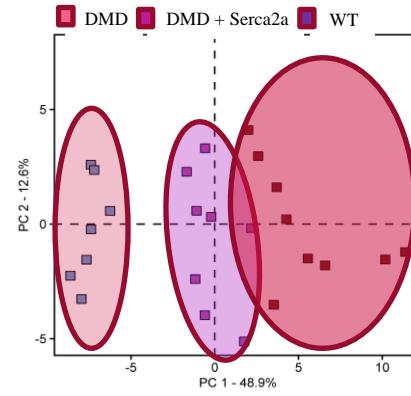
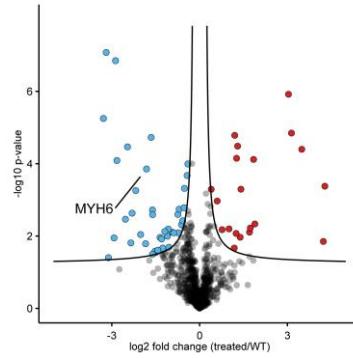


Proteomic analysis of DMD + Serca2a

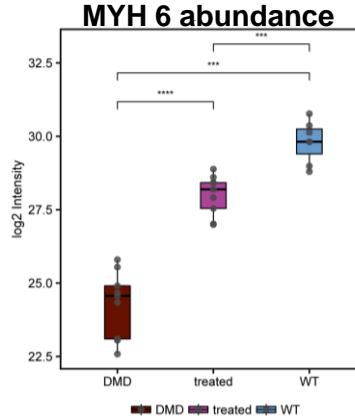
DMD untreated



DMD + AAV.Serca2a



Blue significantly less abundant proteins
Red significantly higher abundant proteins

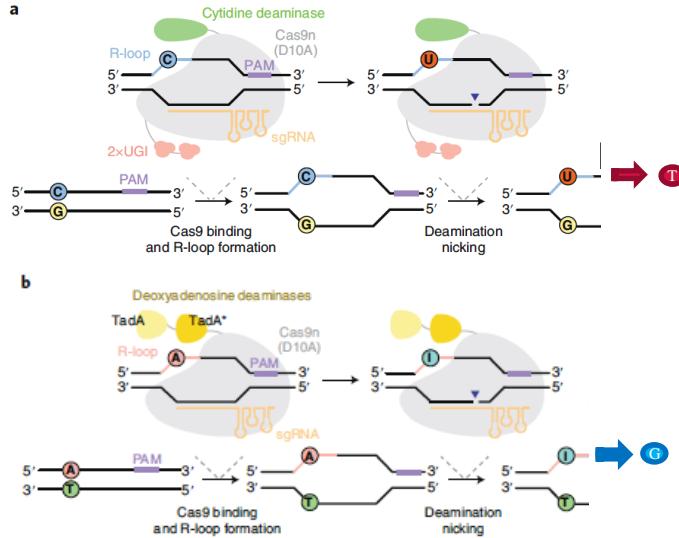


Summary

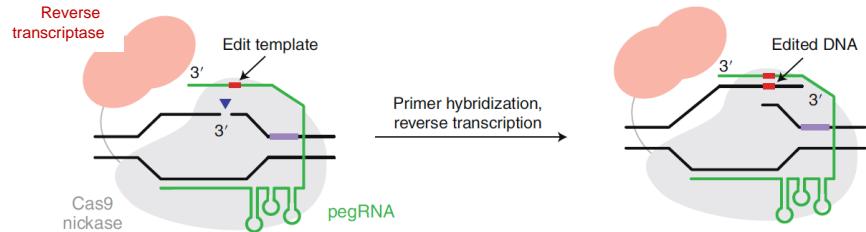
- DMD pigs suffer from heart failure and arrhythmogenic phenotype
- Gene editing can convert DMD to the milder phenotype of BMD, though full cardiac correction is a high aim
- AAV.Serca2a attenuates heart failure and related proteins.
 - Effects on arrhythmias less consolidated

CrispR Cas 2.0 – base and prime editing

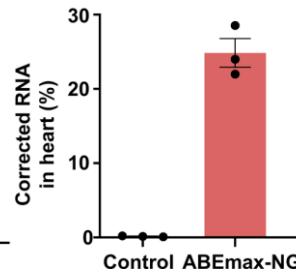
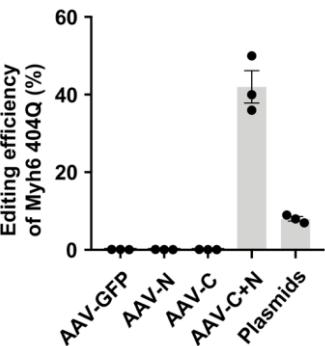
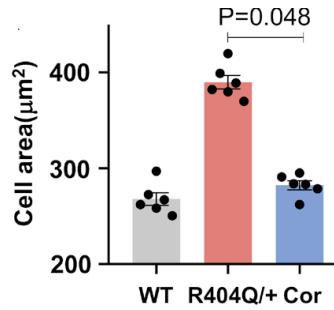
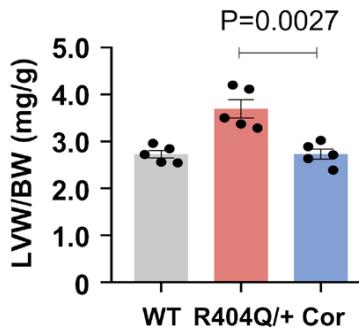
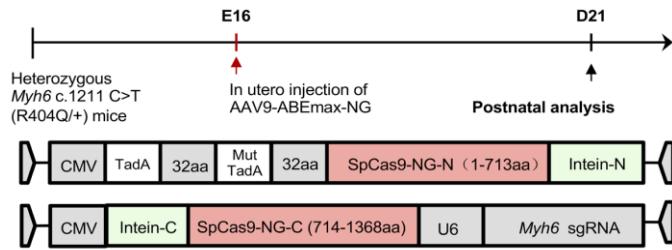
CrispR 2.0 BASE EDITING



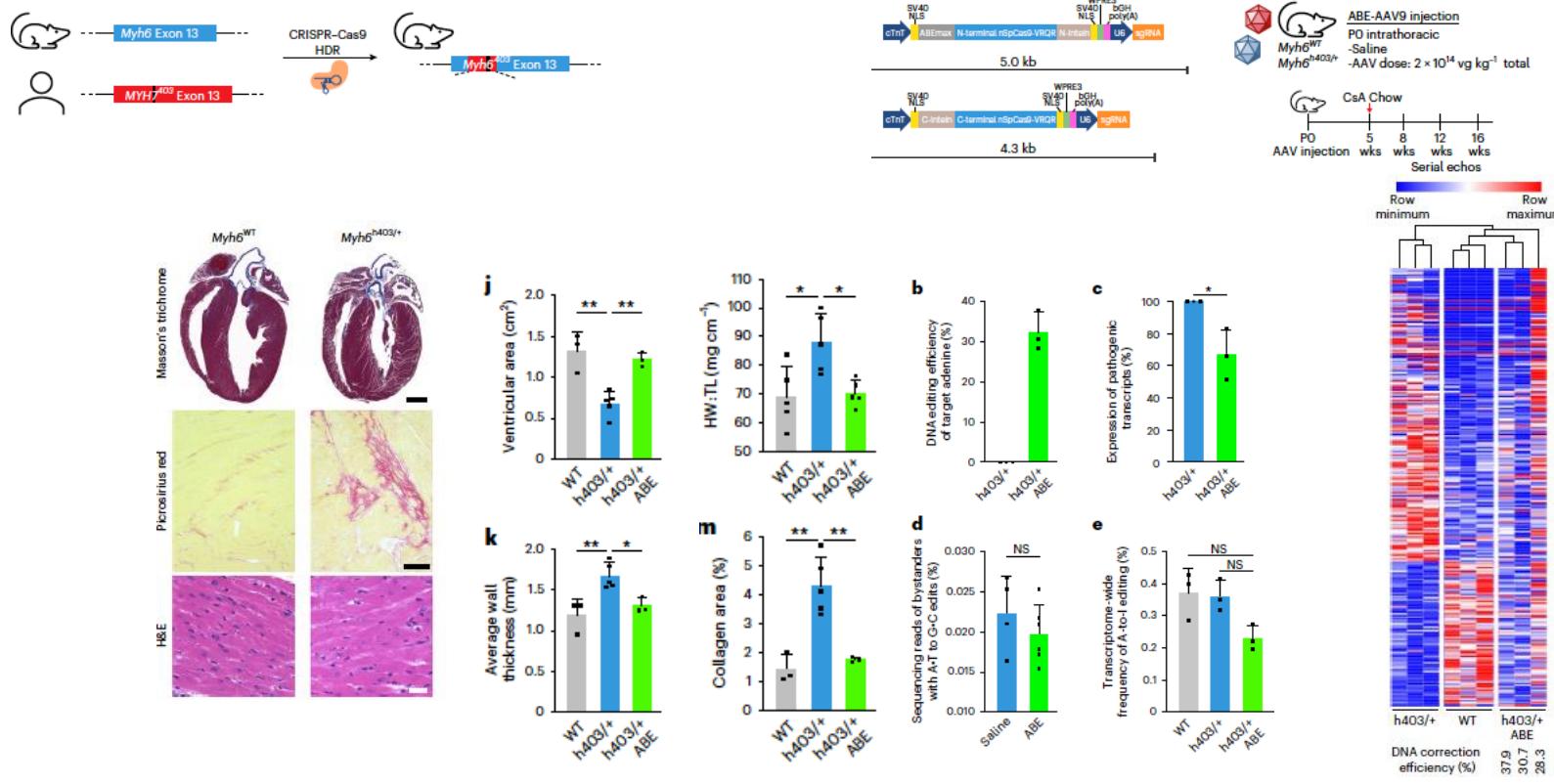
PRIME EDITING



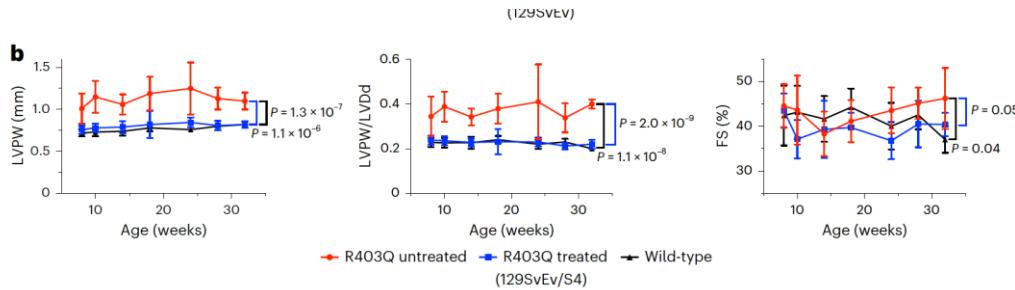
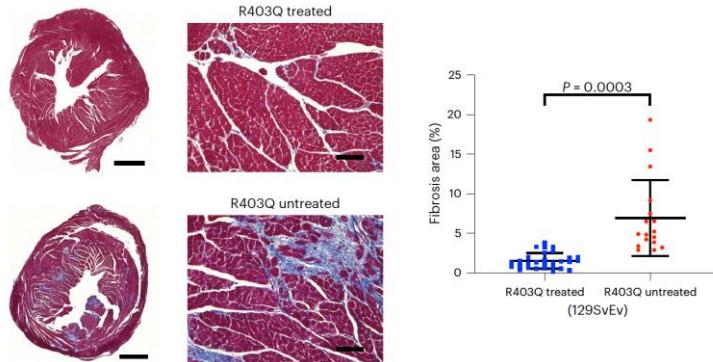
AAV-base-editing of R404Q in embryos



Myosin Heavy Chain Myh6-R403Q base editing



Myosin Heavy Chain 6 R403Q base editing (2)



Summary

- Genetic mutations inducing cardiomyopathies and skeletal muscle dysfunction are increasingly well understood
- This understanding forms the basis for novel therapeutics, either adjuvant or causal (gene editing)
- The presence of genetic cardiomyopathies is at times devastating, the future a lot brighter, and rapidly evolving.

Thanks to

Kupatt Lab (MRI)

Andrea Bähr
Petra Hoppmann
Tarik Bozoglu
Ina Luksch
Anja Wolf
Tilman Ziegler
Nadja Hornaschewitz
Christine Kim
Amelie Hönig

Moretti/Laugwitz Lab (MRI)

Anna B. Meier
Christine Schneider
Daniel Sinnecker
Ralf Dirschinger
Gianluca Santamaria
Tatjana Dorn

AskBio

Roger Hajjar

Wurst Lab (Helmholtz)

Florian Giesert
Jefferey Truong

Collaborators

Manfred Ogris
Remco Megens
Angelika Schnieke
Andreas Dendorfer

Wolf Lab (LMU)

Nik Klymiuk (now own lab)
Michael Stirm
Lina Fonteyne
Barbara Kessler
Mayuko Kurome
Valeri Zhacharchenko

LaFuga
Thomas Fröhlich
Bachuki Shashikadze
Jan Stöckl

